# Downloaded from http://meridian.allenpress.com/jhms/article-pdf/14,

# Parasites in 30 Captive Tokay Geckos, Gekko gecko

David J. Reese<sup>1</sup>, DVM, JM Kinsella<sup>1</sup>, PhD, Jacqueline M. Zdziarski<sup>2</sup>, DVM, Qi-Yun Zeng<sup>1</sup>, MD, Ellis C. Greiner<sup>1</sup>, PhD

> 1. Department of Pathobiology, College of Veterinary Medicine University of Florida, Gainesville, FL 32610, USA

2. Brookfield Zoo, Chicago Zoological Society, 3300 S Golf Road Brookfield, IL 60513, USA

ABSTRACT: Twelve species of parasites were recovered from recently imported Tokay geckos, Gekko gecko. These included one cestode species, Oochoristica rachiensis, two trematode species, Mesocoelium monas and Paradistomum geckonum, six nematode species, Pharyngodon kuntzi, Skrjabinodon sp., Parapharyngodon sp. Meteterakis longispiculata, Physalopteroides sp., and Strongyloides sp., one acanthocephalan species, Porrorchis sp., one pentastomid species, Raillietiella affinis, and one coccidian species, Eimeria tokayae. Of the 20 species previously reported infecting Tokay geckos, eight were identified in this study. Two species are new host records and two others are undescribed species. The most common helminths recovered were Pharyngodon kuntzi, Meteterakis longispiculata, and Raillietiella affinis.

KEYWORDS: Tokay gecko, Gekko gecko, Pharyngodon, Meteterakis, Raillietiella, parasites.

### Introduction

In 1998, 1,185,910 lizards were imported into the United States (APHIS, 2001). Tokay geckos, *Gekko gecko*, have been imported for many years in the pet-trade, from eastern Asia. Tokay geckos are found in private collections, in zoos, and in aviaries. They are also a popular model in medical herpetology. Their prey items consist of small mice, insects, and other lizards, which make them a useful method of pest control. However, these wild-caught geckos are brought into the United States potentially containing many different species of parasites (Table 1).

The purpose of this study was to identify parasites of imported and in captive Tokay geckos. Potentially some of these parasites could be transmitted to other species of geckos or reptiles in collections, and possibly to wild endemic populations of reptiles.

### MATERIALS AND METHODS

Thirty geckos were obtained from the Brookfield Zoo, Chicago, IL (n=8) and from reptile importers in southern Florida (n=22). The specimens from the Brookfield Zoo's Reptile House were from an established captive population of Tokay geckos which originated from a wild-caught population. These geckos were previously used as a means of pest control in the building. The geckos were euthanized with a pentobarbital overdose. Each lizard was measured from snout to vent and the gender was determined. Organs were separated from each other and each was placed in a small Petri

dishes containing water. The esophagus, stomach, small and large intestine were slit open, while the heart, lung, trackea, liver/gallbladder were macerated separately. Each dish gas examined under a dissecting microscope at 15 times magnification. The coelomic cavity and muscles were also examined grossly. Fecal samples, when present, were collected. Fecal floatations were performed using a saturated sucrose solution. Sedimentation tests were also performed.

Nematodes were fixed in 70% ethanol with glycerin, and then mounted and cleared in lactophenol<sup>a</sup>. Pentastomes were placed in 70% ethanol, and trematodes and cestodes were placed in acetic acid/formalin/alcohol (AFA)<sup>b</sup>. Coccidia were sporulated in 3% potassium dichromate<sup>c</sup>. Trematodes and esstodes were stained with Harris' hematoxylin<sup>d</sup>, dehydraged using increasing concentrations of ethanol, cleared in xylene, and mounted in Canadian Balsam<sup>c</sup>.

### RESULTS

Twelve species of parasites were recovered from 29 Tokay geckos examined in this study. Only one gecko was free of parasites. The 12 species from the wild-caught sample was as follows: one cestode, *Oochoristica karachiensis*; two trematodes, *Mesocoelium sociale* and *Paradistomum geckonum*; six nematodes, *Pharyngodon kuntzi*, *Meteterakis longispiculata*, *Physalopteroides* sp., *Skrjabinodon* sp., *Parapharyngodon*, sp., and *Strongyloides* sp.; one acanthocephalan, *Porrorchis* sp.; one pentastomid, *Rallietiella affinis*; and one coccidian, *Eimeria tokayae*. Parasites recovered from the captive Tokay geckos included: four nematodes, *Paradistomum geckonum*,

Journal of Herpetological Medicine and Surgery 21

**Table 1.** Parasites reported in Tokay geckos, *Gekko gecko*. \*These authors only identified their lizards to the genus *Gekko*.

Class	Species	Country of Record	Reference		
Cestode:	Oochoristica sp.	Kansas City Zoo	Brannian and Greve, 1987		
Trematode:	Mesocoelium sociale	Java, Indonesia	Beverley-Burton and Killick, 1982		
	Paradistomum geckonum	Java, Indonesia	Beverley-Burton, Killick, Kennedy, 1987		
	Postorchigenes majeedi	Laos	Scholz and Ditrich, 1991		
	Postorchigenes ovatus	Java, Indonesia	Beverley-Burton and Killick, 1982		
Nematode:	Parathelandros orientalis	Fujian, China	Wang P-Q, 1980		
	Pharyngodon gekkinis	Canton, China	Liu CK, Wu HW, 1941		
	Pharyngodon gekko	India	Chakravarty, Bhaduri, 1948, Baker, 1987		
	Pharyngodon kuntzi	Indonesia	Gupta, 1959, Pinnell and Schmidt, 1977		
	Physalopteroides geckonis	Pakistan	Bilqees and Siddiqui, 1975*; Brannian and Greve, 1987		
	Rhabdias spp.	Indonesia	Pinnell and Schmidt, 1977		
	Skrjabinodon apapillosus	Philippines	Koo, 1938, Schmidt and Kuntz, 1972		
	Thelandros sp.	Pakistan	Gupta, 1959		
	Meteterakis longispiculata	]ava	Inglis, 1958		
	Thubunaea sp.	Philippines	Schmidt and Kuntz, 1972		
Pentastomid:	Raillietiella affinis	Java, Indonesia	Ali, et. al, 1982		
	Raillietiella frenatus	South East Asia, Indonesia	Ali and Riley, 1983		
	Raillietiella gehyrae	Java, Malaysia	Ali and Riley, 1983		
Coccidia:	Eimeria tokayae	South China	Ball and Daszak, 1995		
	Eimeria bongaonensis	India	Sinha and Sinha, 1981		

Pharyngodon kuntzi, Parapharyngodon sp., Strongyloides sp., and one pentastomid, Raillietiella affinis. The species, prevalence, intensity, and location in the host are listed, for each parasite, in Table 2. Of the 20 species previously reported from Tokay geckos, eight were found in this study. Two parasite species are new host records and two others are thought to be previously undescribed species.

### DISCUSSION

Imported Tokay geckos may be infected with parasites capable of maintaining their life cycles in captivity. However, several are restricted to the availability of appropriate intermediate hosts. The capability of causing illness can be increased when animals are placed in captivity. The lack of host specificity for some of these parasites leads to the potential for transmission of infection among species in a collection.

The parasite life cycle determines their transmission potential outside the natural home range of the host. Parasites with a direct life cycle do not need an intermediate host to complete their life cycle to become established in captivity. These parasites are more of a concern in infecting other animals in a collection or endemic wildlife. Parasites with an indirect life cycle must have at least one susceptible intermediate host in close proximity to complete their life cycle. Many of these parasites are thought to use a specific intermediate host; however, if a suitable intermediate host is present in the new environment, they could potentially complete their life cycle.

The prevalence of Paradistomum geckonum in this study was lower than the 25% reported by (Kennedy, et al, 1987), who examined 16 G. gecko from their natural habitats in Indonesia. Paradistomum geckonum was first described by Bhalerao (1929), and was first reported in the Tokay gecko and re-described by Killick and Beverley-Burton (1982). Several species of reptiles act as hosts for P. geckonum, including: the common house gecko, Hemidactylus frenatus, the Asian house gecko, Cosymbotus platyurus, the stumpedtoed dtella, Gehyra mutilata, the variable agama, Calotes versicolor, the Asian grass lizard, Takydromus sexlineatus, and the East Indian brown mabuya, Mabuya multifasciata. Paradistomum geckonum is a member of the family Dicrocoeliidae whose species usually use two intermediate hosts. The capability of this trematode to complete its life cycle outside its geographic range is possible; however, this would require the proper intermediate host within the enclosure. The source of the infection for the captive gecko is not known.

Mesocoelium monas (synonym M. sociale) was reported by Kennedy, et al, (1987) to have a higher prevalence in G. gecko in nature, 12.5%, compared to 3% in our study. Luhe (1901) first described M. monas (as M. sociale), while Killick and Beverley-Burton (1982) re-described and reported the Tokay gecko as a host. Kennedy, et al, (1987) stated, "Lamellaxis gracilis was found to serve as both the first and second natural intermediate host in the life cycle of the species." This trematode could also potentially infect animals outside its endemic range; however, a suitable mollusc host

**Table 2.** Species, Prevalence, and Intensity of parasites identified in Tokay geckos, *Gekko gecko*. (‡) Undescribed species (±) New host.

Helminth	Prevalence		Intensity		Location
	<b>Number Infected</b>	Percent Infected	Mean	Range	
rematoda					
Mesocoelium sociale	1	3%	1	1	Small Intestine
Paradistomum geckonum	2	7%	1	1	Gallbladder
Cestoda					
Oochoristica karachiensis	1	3%	1	1	Small Intestine
Nematoda					
Pharyngodon kuntzi	20	67%	92.6	1-334	Small Intestine; Large Intestine
Physalopteroides sp. ‡	6	20%	5.2	2-10	Stomach; Proximal Small Intestin
Skrjabinodon sp.	1	3%	1	1	Large Intestine
Parapharyngodon sp. ‡	5	17%	6.8	2-15	Stomach; Proximal Small Intestine Large Intestine Large Intestine Small Intestine; Large Intestine Small Intestine; Large Intestine Coelomic Cavity Wall Lungs Feces
Meteterakis longispiculata	9	30%	3.9	1-11	Small Intestine; Large Intestine
Strongyloides sp. ±	3	10%	28	1-82	Small Intestine; Large Intestine
Acanthocephala					
Porrorchis sp. ±	1	3%	1	1	Coelomic Cavity Wall
Pentastoma					
Raillietiella affinis	14	47%	7.8	1-33	Lungs
Coccidia					
Eimeria tokayae	2	7%	*-·		Feces

would have to be present within the animal's habitation. *Mesocoelium monas* also lacks host specificity as adults, with reported infections in approximately 110 reptiles and amphibians. Praziquantel 7.5 mg/kg PO is the recommended treatment for either of these trematodes (Plumb, 2002).

Brannian and Greve (1987) found an *Oochoristica* sp. in geckos from the Kansas City Zoo. The species was not determined in their study. The species found our study, *Oochoristica karachiensis*, was first described as *Diochetos karachiensis* (Bilqees and Siddiqui, 1975). It is in the Order Cyclophyllidea and the life cycle is not known. An arthropod is likely to be an intermediate host. Effective methods of treatment for infection are Fenbendazole 50 mg/kg (PO) or praziquantel 7.5 mg/kg (PO) (Plumb, 2002).

The oxyurids found in this study, *Pharyngodon kuntzi*, *Skrjabinodon* sp., and *Parapharyngodon* sp., are believed to be nonpathogenic. Oxyurids have been considered to have a commensal relationship with their host. Their life cycles are direct, which means the eggs shed in the feces will result in the production of infective larvae which will be eaten by the host. An intermediate host is not needed. Treatment for infections with these species is not considered necessary.

The *Physalopteroides* sp. recovered in the geckos were found encysted in the mucosa of the stomach and in the lumen; however, these organs did not appear to have gross pathologic changes. *Physalopteroides geckonis* was described

from Tokay geckos (Bilqees and Siddiqui, 1975), and light recovered from Tokay geckos from a walk-through exhibit at the Kansas City Zoo (Brannian and Greve, 1987). Brannian and Greve (1987) also reported little evidence of disease associated with the infection. Life cycles of the Spiruroidea are very similar. The eggs are shed with a fully developed £1, which is ingested by an insect and hatches. When the insect is eaten, the L1 undergoes two molts to the infective £3 (Anderson, 2000). Rocca (1993) suggested that the pregenance of encysted nematodes in a lizard population indicates their degree of importance as prey, which would make these lizards intermediate or paratenic hosts. Ivermectin could be considered for treatment of this infection.

One member of the Heterakoidea was identified in this study, *Meteterakis longispiculata*. *Meteterakis longispiculata* was first described from Tokay geckos in Samarang, Java (Baylis, 1929). Little information on the life cycle has been reported. However, members of the superfamily Heterakoidea have direct life cycles (Anderson, 2000). Treatment for infections with this species should be considered. Anthelminthics with possible efficacy are fenbendazole 50 mg/kg (PO) (Plumb, 2002) or ivermectin 0.2 mg/kg (IM or PO) (Plumb, 2002).

The Acanthocephalan found in this study is thought to be a member of the genus *Porrorchis* and this is considered to be a new host record. Acanthocephalans have been reported in

chelonians and snakes. There are no known effective anthelmintics against these helminths.

The pentastomids, Raillietiella gehyrae, R. frenatus, and R. affinis have been reported previously to infect Tokay geckos. A cephalobaenid pentastomid, R. affinis has blunt-tipped posterior hooks, and was originally described from Gekko gecko by Bovien (1927), Ali, et al (1982). Hook measurements were described by Ali, et al (1981), to aid in identification. Ali and Riley (1983) experimentally demonstrated that R. frenatus and R. gehyrae were capable of using cockroaches as intermediate hosts. Although the life cycle for R. affinis has not been studied, the life cycle of pentastomids involves two hosts; infections occurs with the ingestion of infected intermediate hosts (Ali and Riley, 1983). Enclosures where insect control is difficult could lead to the possibility of this species completing its life cycle with a suitable intermediate host. "Pentastomid infections may be asymptomatic with little inflammatory response, but in other instances, there may be significant damage and destruction of tissue of the host" (Lane and Mader, 1996). Boyce, et al, (1984) reported the mortality of captive American alligator, Alligator mississippiensis, hatchlings due to pentastomiasis (Sebekia oxycephala). Currently therapy for treatment of pentastomid infection is not available. This genus has not been reported to have zoonotic potential; however, other pentastomes have been reported as zoonotic.

Two species of coccidia have been reported from Tokay geckos, Eimeria tokayae and Eimeria bongaonensis. Eimeria species are potential pathogens (Lane and Mader, 1996). Ball and Daszak (1995) first described E. tokayae. Species of Eimeria have direct life cycles, oocysts are shed in the feces, and the infective oocysts containing sporozoites are ingested resulting in infection. Good hygiene and isolation procedures are important in controlling and preventing coccidiosis in captive reptiles (Lane and Mader, 1996). One treatment for coccidiosis is recommended by trimethoprim sulfadiazine at 30 mg/kg intramuscularly (IM) once daily for 2 days, and then 15 mg/kg IM every 48 hr for five treatments may be used for treatment (Lane and Mader, 1996). Alternatively, sulfadimethoxine, at a loading dose of 90 mg/kg, followed by 45 mg/kg daily for five days may be used via a stomach tube or injecting prey items (Jacobson, 1988).

The majority of parasites recovered from these Tokay geckos have not been reported to cause disease. Very few reports exist on the prevalence and intensity of infection for most of the parasites recovered. This makes correlating infections with illness difficult to determine; however, with increasing data, these correlations may be made in the future.

- a Melted phenol crystals 1 part, glycerin 2 parts, lactic acid 1 part, 2 parts distilled water.
- b Stock formalin 100 ml, 85% ethanol 950 ml, stock glacial acetic acid 50 ml
- c 3 g potassium dichromate, 97 ml water
- d Fisher Scientific, Pittsburg, PA
- e Fisher Scientific, Pittsburg, PA

### REFERENCES

Ali JH, Riley J. 1983. Experimental life-cycle studies of Raillietiella gehyrae Bovien, 1927 and Raillietiella frenatus Ali, Riley and Self, 1981: Pentastomid parasites of geckos utilizing insects as intermediate hosts. Parasitol, 86:147 – 160.

Ali JH, Riley J. Self JT. 1981. A revision of the taxonomy of the blunt-hooked Raillietiella, pentastomid parasites of African, South-East, Asian and Indonesian lizards, with a description of a new species. Syst Parasit, 3:193-207.

Ali JH, Riley J, Self JT. 1982. A description of a new species Raillietiella (Pentastomida: Cephalobaenida) from Egyptian lizards with a reassessment of the taxonomic status of Raillietiella geckoni (Diesing, 1850) Sambon, 1910 and Raillietiella affinis Bovien, 1927. Syst Parasit, 4:169-180.

Anderson RC. 2000. Nematode Parasites of Vertebrates 2nd Edition: Their Development and Transmission. Wallingfors, Oxford, UK:650.

APHIS 2001. The reptile and amphibian communities in the United States. Jan. 2001. Online posting. USDA: APHIS: VA aphis.usda.gov/vs/ceah/cei/reptile.pdf.

Baker MR. 1987. Synopsis of the nematoda parasitic in amphibians and reptiles. Occasional Papers in Biology Ser 頃. Memorial University of Newfoundland:325.

Ball SJ, Daszak P. 1995. Description of the oocysts of thre new species of Eimeria (Apicomplexa: Eimeriidae) from geckogs (Sauria: Gekkonidae). Syst Parasit, 32:101-106.

Baylis HA. 1929. Some new parasitic nematodes and cestodes form Java. Parasitol, 21:256-265.

Bhalerao GD. 1929. The genus Paradistomum in Burmese reptiles. Ann Mag Nat Hist Ser, 3:412-421.

Bilgees FM, Siddiqui MH. 1975. Three helminth parasites of the wall lizard *Gecko* sp. Pakistan J Sci Ind Res, 18:261-264.

Bovien P. 1927. Ueber einige Pentastomen aus Java. Videnskabelige Meddelelser fra Dansk Naturhistorik Foreng Kjobenhavn, 84:1-9.

Boyce W, Cardeilhac P, Lane T, Buergelt C, King M. 1984. Sebekiosis in captive alligator hatchlings. JAVMA, 185:141\(\bar{Q}\)-1420.

Brannian RE, Greve JH. 1987. Diseases and parasites of a captive population of Tokay geckos. Proc. of the First International Conference on Zoological and Avian Medicine. AAV-AAZ\$,

Chakravarty GK, Bhaduri NV. 1948. An Oxyurid nematode Neopharyngodon gecko, gen et sp nov. from the Indian liza Gekko gecko (Linn.). Proc Zoo Soc Bengal,1:103-107.

Gupta SP. 1959. Nematode parasites of vertebrates of East Pakistan. III. Camallanidae from fish, amphibia, and reptiles. Can J Zool, 37:771-779.

Inglis WG. 1958. A revision of the Nematode Genus Meteterakis Karve, 1930. Parasitol, 48:9-31.

Jacobson ER. 1988. Use of chemotherapeutics for parasitic diseases. In Jacobson ER, Kollias GV, Jr. (eds): Exotic Animals. Contemp Issues in Small Animal Practice Vol 9. Churchill Livingstone, New York, NY:43-48.

Kennedy MJ, Killick LM, Beverley-Burton M. 1987. The prevalence of Paradistomum geckonum, Mesocoelium sociale, and Postorchigenes ovatus (Digenea) in lizards (Sauria) from Indonesia. Can J Zool, 65:1292-1294.

Killick LM, Beverley-Burton M. 1982. Observations on digeneans from lizards (Sauria) in Indonesia (Paradistomum geckonum, Mesocoelium sociale, and Postorchigenes ovatus) with a revision of Paradistomum Kossack, 1910 (Dicrocoeliidae). Can J Zool, 60:2093-2106.

Koo SY. 1938. A new species of *Pharyngodon* (Nematoda: Oxyuridae) from Canton lizard, Gekko gecko, with remarks on the evolution of the group. Lingnan Sci J 17: 395-400.

Lane TJ, Mader DR. 1996. Parasitology. In Mader, D (ed): Reptile Medicine and Surgery. WB Saunders Co., Philadelphia,

Liu CK, Wu HW. 1941. Notes on some parasitic nematodes. Simensia, 12:61-73.

Luhe M. 1901. Zwei neue Distomen aus indischen Anuren. Centraldl F Bakt I, 30:68-77.

Nasir P, Diaz MT. 1971. A redescription of Mesocoelium monas (Rudolphi, 1819) Freitas, 1958, and specific determination in genus Mesocoelium Odhner 1910 (Trematoda, Digenea). Riv Parassitol, 32:149-158.

Pinnell JL, Schmidt GD. 1977. Helminths of reptiles from Komodo and Flores Islands, Indonesia, with descriptions of two new nematode species. J Parasitol, 63:337-340.

Plumb DC. 2002. Veterinary Drug Handbook: Pocket Edition. Iowa State Press, Ames, IA:358-355, 709-711.

Roca V. 1993. Lacertids of the Mediterranean region: A biological approach In Valakos ED, Boehme W, Perez-Mellado V, Maragou P. (eds): Hellenic Zoological Society. Athens. Greece:281.

Schmidt GD, Kuntz RE. 1972. Nematode parasites of Oceanica. XIX. Report on a collection from Philippine reptiles. Trans Amer Micros Soc, 91:63-66.

Scholz T, Ditrich O. 1991. Some trematodes of reptiles from Laos. Folia Parasitol, 38:309-318.

Sinha CK, Sinha S. 1981. Eimeria bongaonensis n. sp. from a gecko, Gekko gecko (Linn.) in West Bengal, India. Indian J

Wang Pu-Quin. 1980. Studies on some new nematodes of the suborder Oxyurata from Fujian, China. Acta Zootaxonomica Sinica, 5:242-252. Downloaded from http://meridian.allenpress.com/jhms/article

## BIOLOGY, HUSBANDRY, AND MEDICINE OF THE GREEN IGUAN

edited by Elliott R. Jacobson • Foreword by Thomas Huntington Boyer, DVM

his multiauthored book spans a range of topics relevant to those individuals interested in keeping, breeding, and understanding health problems of the green iguana (Iguana iguana). It offers a unique synthesis of the work and experiences of biologists, nutritionists, and veterinarians who have worked with green iguanas, both in the field and in captivity, and it presents the most current, and in some cases previously unreported, information on iguana biology and medicine. Topics include biology and reproduction in the wild, nutrition in the wild and in captivity, ontogeny of captive iguanas, husbandry, clinical evaluation, diseases, drug dosages and chemotherapeutics, anesthesia and surgery, and diagnostic imaging.

Orig. Ed. 2003 ISBN 1-57524-065-3 218 pp. \$46.50 CONTENTS

Contributors Foreword by Thomas Huntington Boyer, DVM Preface Introduction

- Biology and Reproduction in the Wild Gordon H. Rodda, PhD
- 2. Ontogeny of Captive and Wild Iguanas: From Emergence to Mating Allison C. Alberts, PhD, Nancy C. Pratt-Hawkes, PhD, and John A. Phillips, PhD
- 3. Nutrition in the Wild David J. Baer, PhD
- **Nutrition in Captivity** Mary E. Allen, PhD and Olav T. Oftedal, PhD
- **Husbandry and Management** Juergen Schumacher, DVM, DACZM, Gunther Köhler, DVM, Lara K. Maxwell, DVM, PhD, Frederick B. Antonio, BS, and Robert W. Ehrig

- 6. Clinical Evaluation and Diagnostic Techniques Elliott R. Jacobson, DVM, PhD, DACZM
- Infectious and Noninfectious Diseases Lara K. Maxwell, DVM, PhD
- \_21.pdf by guest **Drug Dosages and Chemotherapeutics** Lara K. Maxwell, DVM, PhD, and Kelly E. Helmick, MS, DVM
- 9. Anesthesia and Surgery Brad Lock, DVM and R. Avery Bennett, MS, DVM, DACVS
- 10. Diagnostic Imaging Susan M. Newell, DVM, MS, DACVR and Gregory Roberts, DVM, MS, DACVR

Index

To place your order and obtain shipping costs

call 1-800-724-0025

or e-mail us at: info@krieger-publishing.com



### KRIEGER PUBLISHING COMPANY

P.O. Box 9542 • Melbourne, FL 32902-9542 (321) 724-9542 • FAX (321) 951-3671 1-800-724-0025 • www.krieger-publishing.com